

## **β Gal-Juice/ β Gal-Juice PLUS (1 033 11/ 1 033 12)**

### **Components included:**

<b>β-Gal-Juice/plus</b>	<b>50ml</b> Chemiluminescent Substrate for measurement of β-Galactosidase Enzyme. <b>Store at +4°C.</b>
<b>Triggering Reagent</b>	<b>50ml</b> <b>Store at +4°C</b>

### **Preparation of Cell Lysates:**

For cell lysis, we suggest the follow buffers:

- **Animal cells:** 100 mM potassium phosphate (pH 7,8), 0,2% Triton X100
- **Yeast cells :** 0,5 M sodium phosphate (pH 7,1) 50 mM KCl, 5 mM MgSO<sub>4</sub>

### **Standard Protocol for Detection of β-Galactosidase in Microtiter Plate Luminometer**

#### **Equilibrate at room temperature for 30 minutes before use.**

Best results for chemiluminescent detection of β-Galactosidase enzyme or reporter assays can be obtained from 30 minutes to 60 minutes incubation of β-Gal Juice with β-Galactosidase enzyme and read the plate or tube within 2 to 10 minutes after triggering the reaction mixture by accelerator or enhancer.

- Add 5 to 10 µl of diluted β-Galactosidase enzyme or cell extract to microplate wells.
- Add 50 to 100 µl of β-Gal Juice and incubate for 30 to 60 minutes at room temperature.
- Add 50 to 100 µl of triggering reagent and shake it for 10 seconds.
- Place the microtiter plate in luminometer and start reading.
- Best results are obtained between 2 to 10 minutes.

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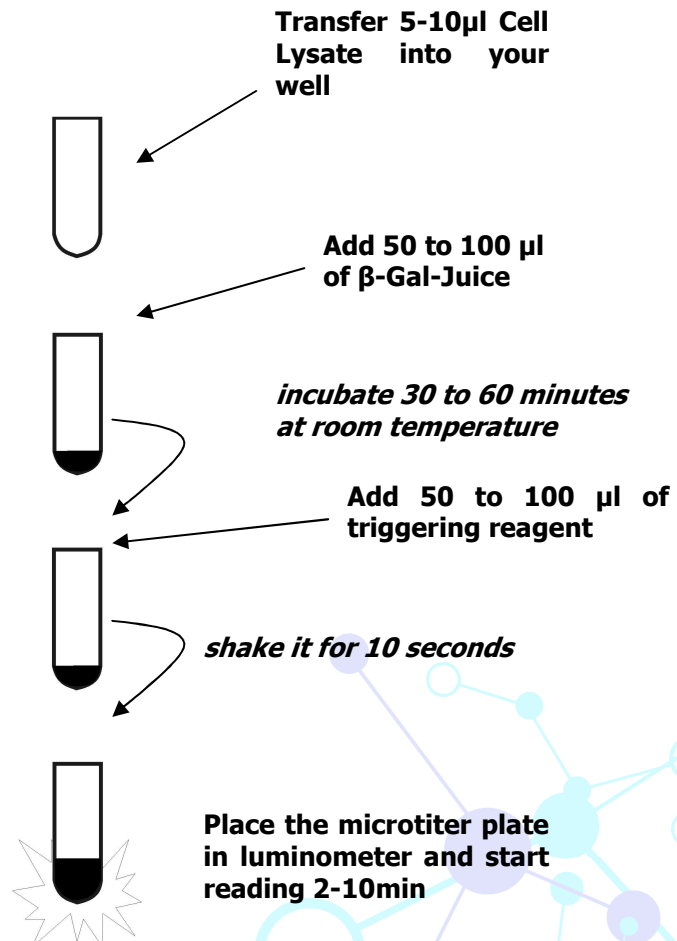
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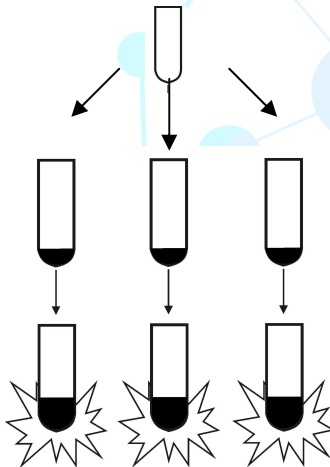
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### Standard procedure:



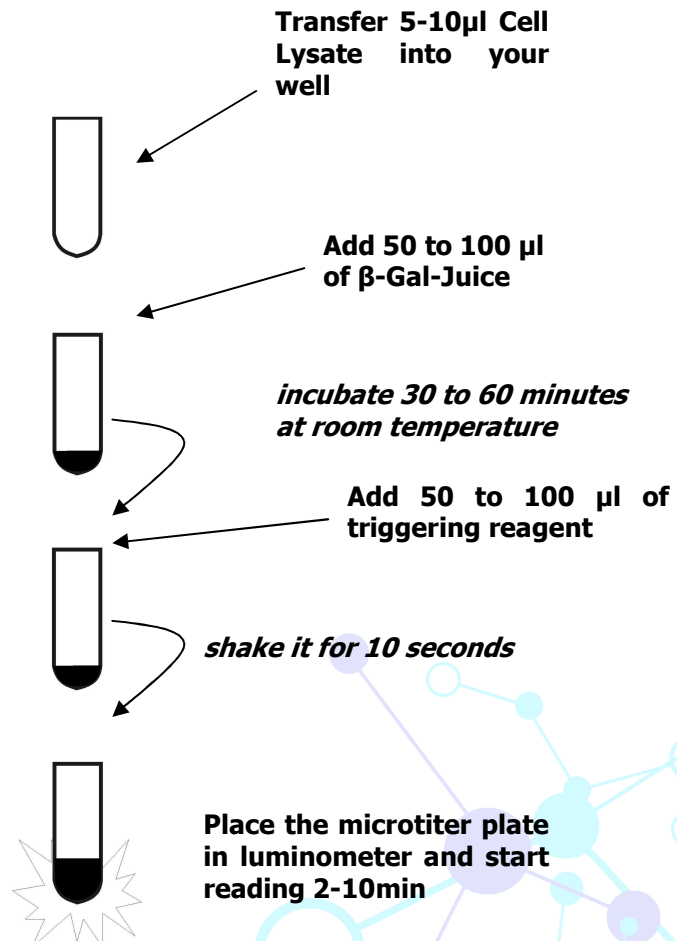
### Optional: parallel measurement in Duo- or Triple-Well System (Firefly/ Renilla/ Gaussia/ AP/ $\beta$ Gal)

Split the cell lysate and transfer to different tubes for measuring several enzymes in one sample.



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